

# ORGANIC COMPOUNDS

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## Three Substituted 3-Amino-2-benzoylamino-propenoic Acid Derivatives

KRISTINA DJNOVIĆ-CARUGO,\*† LJUBO GOLIČ,  
IVAN LEBAN, JURIJ SVETE, BRANKO STANOVNÍK  
AND MIHA TIŠLER

*Department of Chemistry and Chemical Technology,  
University of Ljubljana, Murnikova 6, Ljubljana 61001,  
Slovenija*

CECIL TATE

*Physics Department, University of York,  
York Y01 5DD, England*

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### Abstract

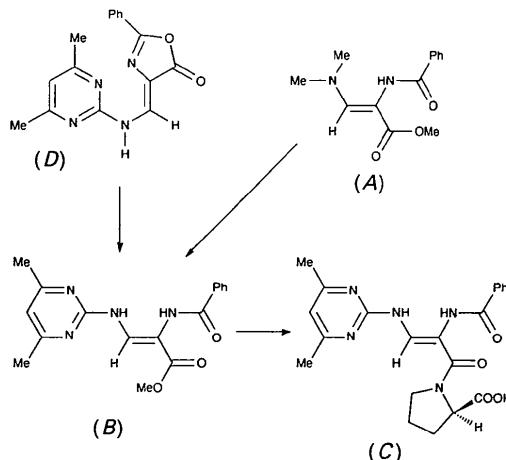
The molecular structures of three substituted 3-amino-2-benzoylamino-propenoic acid derivatives, methyl (*Z*)-2-benzoylamino-3-dimethylaminopropenoate (*A*), methyl (*Z*)-2-benzoylamino-3-(4,6-dimethyl-2-pyrimidinylamino)propenoate (*B*) and *N*-[*(Z*)-2-benzoylamino-3-(4,6-dimethyl-2-pyrimidinylamino)propenoyl]-L-proline monohydrate (*C*), show that the configuration around the exocyclic C=C double bond is *Z*. The mechanisms of syntheses of (*B*) and (*C*) from (*A*) are confirmed as the *Z* configuration is preserved in the title compounds. The crystal structure of compound (*C*) is stabilized via hydrogen bonds from the solvent water molecules to the N atom bonded to the C=C double bond and to the two carbonyl O atoms in the molecule, each of the water molecules contributing to three hydrogen bonds. The bond lengths and angles agree with expected values.

### Comment

Dehydroamino acids are of major interest in research on bioactive dehydropeptides and asymmetric hydrogenation (Schmidt, Häusler, Öhler & Poisel, 1979; Stammer, 1982; Kagan, 1985; El-Baba, Nuzillard, Paulin & Kagan, 1986; Schmidt, Lieberknecht & Wild, 1988).

† Current address: Dipartimento di Genetica e Microbiologia, Sezione di Biologia Molecolare e Biofisica, Università di Pavia, Via Abbiategrasso 207, I-27100 Pavia, Italy.

Methyl (*Z*)-2-benzoylamino-3-dimethylaminopropenoate (*A*), prepared either in a two-step (Japan Kokai, 1975) or in a one-step synthesis from hippuric acid (Stanovník, Sveté, Tišler, Žorž, Hvala & Simonič, 1988), has been introduced recently as a versatile reagent for the synthesis of  $\beta$ -arylamino- $\alpha,\beta$ -dehydro- $\alpha$ -amino acid derivatives (Stanovník, Urbanija, Sveté & Tišler, 1989),  $\beta$ -heteroaryl amino- $\alpha,\beta$ -dehydro- $\alpha$ -amino acids and dipeptides, such as (*B*) and (*C*) (Stanovník *et al.*, 1988), as an alternative to the method where oxazolones, such as (*D*), are formed as intermediates (Stanovník, Sveté & Tišler, 1987; Sveté, Stanovník, Tišler, Golič & Leban, 1989), and monocyclic, bicyclic and polycyclic systems, where the  $\alpha$ -amino acid structural element is incorporated into the ring systems, such as pyranones (Sveté, Čadež, Stanovník & Tišler, 1990), benzopyranones (Stanovník, Sveté & Tišler, 1989), pyranobenzopyranones (Ornik, Čadež, Stanovník & Tišler, 1990), isomeric naphthopyranones and naphthodipyranoines (Ornik, Stanovník & Tišler, 1992a,b), pyranoazoles and pyranoazines (Stanovník *et al.*, 1989; Stanovník, Golič, Kmecl, Ornik, Sveté & Tišler, 1991), including pyranoquinoline derivatives (Kmetič, Stanovník & Tišler, 1993), and azolo- and azinopyrimidines (Stanovník, van de Bovenkamp, Sveté, Hvala, Simonič & Tišler, 1989).



X-ray structure analysis of (*D*) (Leban, Sveté, Stanovník & Tišler, 1991) showed that the configuration of the heteroaryl amino group and the N atom of the oxazolinone ring is *Z* around the exocyclic C=C double bond. The crystal structure determination was undertaken to elucidate the molecular structures and synthetic mechanisms of the title compounds.

Views of the molecules in the same orientation along with the atomic numbering are shown in Figs. 1–3. The observed bond distances and angles are within the expected values when compared to those reported in *Tables of Bond Lengths Determined by X-ray and Neutron Diffraction*, Part I (Allen, Kennard, Watson, Brammer, Orpen & Taylor, 1987).

The observed configuration of N atoms around the C=C bond is Z in all three compounds. The C=C bond distances in compounds (A) and (B) are somewhat longer [1.350 (5) and 1.332 (3) Å, respectively] when compared to the same distances of the two molecules of compound (C) in the asymmetric unit [1.325 (5) and 1.317 (5) Å for (I) and (II),

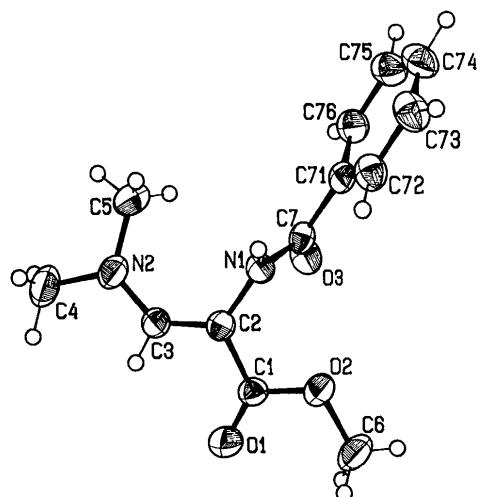


Fig. 1. ORTEP diagram (Johnson, 1965) showing the molecular structure and atom-labelling system of molecule (A). Non-H atoms are shown as 50% probability ellipsoids.

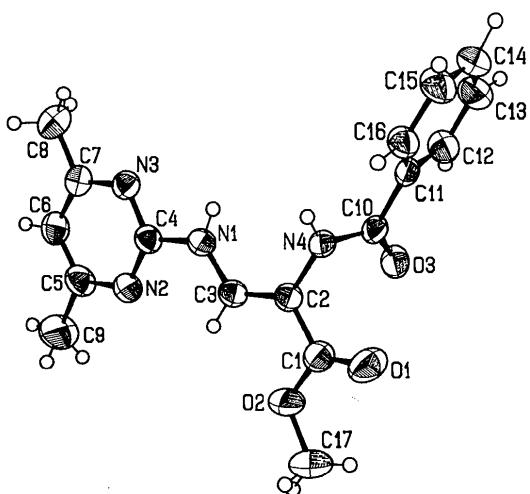


Fig. 2. ORTEP diagram (Johnson, 1965) showing the molecular structure and atom-labelling system of molecule (B). Non-H atoms are shown as 50% probability ellipsoids.

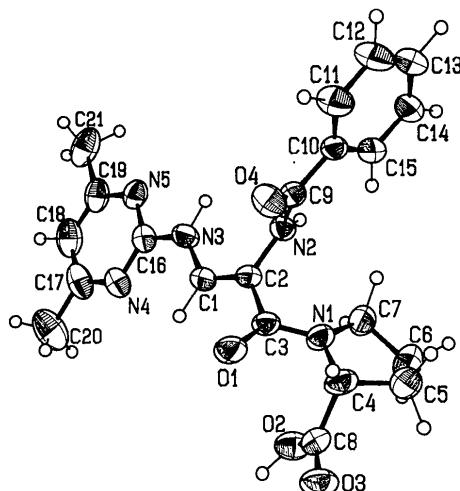


Fig. 3. ORTEP diagram (Johnson, 1965) showing the molecular structure and atom-labelling system of molecule (I) of the asymmetric unit of (C). Non-H atoms are shown as 50% probability ellipsoids.

respectively]. The geometric parameters of the phenyl rings in all four molecules can be considered to be equal within the limits of experimental error. While in compounds (A) and (B) the carbonyl groups adjacent to the phenyl ring are oriented under the plane defined by the N—C=C—N moiety, free rotation around the C—N bond in compound (C) is ‘clipped’ at the point where the carbonyl group is above this plane, as seen Figs. 1–3. This conformation might be explained by hydrogen-bonding interactions of carbonyl atoms O(1) and O(4) with O atoms of the water molecules O(10) and O(20) [for molecules (I) and (II) of the asymmetric unit, see Table 7], stabilizing the molecular structure. The influence of this hydrogen bond is also reflected by the carbonyl C—O bond distance which is longer in compound (C) [1.235 (4) and 1.230 (4) Å, C(9)—O(4)] with respect to the bond lengths in compounds (A) and (B) [1.226 (4) Å, C(7)—O(3) and 1.231 (2) Å, C(10)—O(3)], respectively. The same carbonyl O atom seems also to have a structural role in compound (B) where it is involved in hydrogen-bonding interactions with the N atoms around the C=C double bond of symmetry-related molecules. In the crystal structure of (C) the solvent molecules also take part in hydrogen bonding to the peptide N atoms N(2): O(10) interacts with the N(2) atom of molecule (II), while O(20) is within hydrogen-bonding distance of N(2) of the symmetrically related molecule (I), contributing to the stability of the crystal structure of compound (C).

The energy difference between *cis* and *trans* isomers in di-, tri- and polypeptide chains is about 8.4 kJ mol<sup>-1</sup>, the *trans* isomer having the lower energy (Ramachandran, Ramakrishnan & Sasisek-

haran, 1963). In proline the difference is smaller due to the reduction in the contact between  $C^\alpha$  and  $C^{\delta,\gamma}$  during the transformation from *cis* to *trans*. This is the reason for the occurrence of *cis* bonds in small cyclic peptides at the N-terminal end of proline (Wieland & Birr, 1976), as well as in some isolated cases in globular protein structures (Epp, Lattman, Schiffer, Huber & Palm, 1975). In the structure of (C), the proline is found to have the *trans* conformation. The angle  $C(2)—C(3)—N(1)$  is 118.2 (3) and 117.9 (3) $^\circ$  for molecules (I) and (II), respectively, of the asymmetric unit, agreeing with the normal value of 118 $^\circ$  (Pauling, 1960). The angle  $O(1)—C(3)—N(1)$  is 121.8 (3) and 120.8 (3) $^\circ$  in (I) and (II), respectively, comparable to the mean value of 120.3 $^\circ$  found in some oligopeptides (Ashida & Kakudo, 1974).

Proline rings can also be divided in two classes with respect to the torsion angle  $\chi_1$ . In class A the torsion angle  $\chi_1$  takes negative values, while in class B the values are positive (Balasubramanian, Lakshminarayanan, Sabesan, Tegoni, Venkatesan & Ramachandran, 1971). In compound (C) the proline ring has the B conformation with the  $\chi_1$  torsion angles defined by  $N(1)—C(4)—C(5)—C(6)$  of 28.6 (5) and 21.6 (6) $^\circ$  for molecules (I) and (II), respectively.

The rotations around the  $N—C^\alpha$  and  $C^\alpha—C$  bonds of the peptide linkage are denoted by  $\varphi$  and  $\psi$  torsional angles (Ramachandran & Ramakrishnan, 1965). In our compounds the  $C^\alpha$  atom is  $sp^2$  hybridized and the torsion angles for compound (C) are:  $\varphi = -62.7$  (4),  $-73.4$  (4) $^\circ$  [ $C(9)—N(2)—C(2)—C(3)$ ];  $\psi = 39.6$  (4),  $44.6$  (4) $^\circ$  [ $N(2)—C(2)—C(3)—N(1)$ ] for the two molecules in the asymmetric unit. The corresponding values of  $\varphi$  in compounds (A) and (B) are 75.7 (5) [ $C(7)—N(1)—C(2)—C(1)$ ] and 62.3 (3) $^\circ$  [ $C(10)—N(4)—C(2)—C(1)$ ], respectively, reflecting again the different orientation of the carbonyl group linked to the phenyl ring.

The peptide bond is in *trans* conformation in all three compounds, with the  $\omega$  angles ranging from 173.9 (3) to 179.8 (2) $^\circ$ . The same *trans* conformation is also evident in the second peptide linkage of compound (C), where the  $\omega$  angles [ $C(10)—C(9)—N(2)—C(2)$ ] are 173.6 (3) and 176.0 (3) $^\circ$  for (I) and (II), respectively. The  $\varphi$  angles for the terminal proline residue of compound (C) [ $C(3)—N(1)—C(4)—C(8)$ ] are 62.8 (4) and 58.1 (4) $^\circ$  for (I) and (II), respectively.

The orientation of the substituents around the  $C=C$  double bond in the three compounds is similar. The two C atoms and the two bonded N atoms are nearly coplanar with the heterocyclic ring in compounds (B) and (C); the heterocyclic ring plane and the  $N—C=C—N$  plane form angles of 10.1 (1) $^\circ$  for (B), and 7.9 (1) and 8.0 (1) $^\circ$  for (C); the dimethylamino group of compound (A), defined by

atoms  $N(2)$ ,  $C(4)$  and  $C(5)$ , forms an angle of 9.0 (4) $^\circ$  with the  $N—C=C—N$  plane. Similarly, the phenyl rings in (A), (B) and (C) form angles of 65.3 (2), 83.7 (1), 90.2 (2) and 105.3 (2) $^\circ$  with the  $N—C=C—N$  plane, respectively, while the pyrrolidine ring forms angles of 51.7 (2) and 53.7 (2) $^\circ$  with this plane in the two molecules of the asymmetric unit of compound (C).

## Experimental

### Compound (A)

#### Crystal data

$C_{13}H_{16}N_2O_3$	Mo $K\alpha$ radiation
$M_r = 248.28$	$\lambda = 0.71069 \text{ \AA}$
Orthorhombic	Cell parameters from 50 reflections
$Fdd2$	$\theta = 6.16\text{--}12.51^\circ$
$a = 41.505$ (6) $\text{\AA}$	$\mu = 0.866 \text{ mm}^{-1}$
$b = 19.450$ (2) $\text{\AA}$	$T = 293 \text{ K}$
$c = 6.338$ (1) $\text{\AA}$	Prismatic
$V = 5117$ (2) $\text{\AA}^3$	$0.40 \times 0.24 \times 0.08 \text{ mm}$
$Z = 16$	Colourless
$D_x = 1.289 \text{ Mg m}^{-3}$	

#### Data collection

Enraf-Nonius CAD-4	$R_{\text{int}} = 0.040$
diffractometer	$\theta_{\text{max}} = 28^\circ$
$\omega$ - $2\theta$ scans	$h = 0 \rightarrow 54$
Absorption correction:	$k = -25 \rightarrow 25$
none	$l = -8 \rightarrow 8$
6231 measured reflections	3 standard reflections
1676 independent reflections	frequency: 125 min
822 observed reflections	intensity variation: 1.8%
$ I  > 2.5\sigma(I)$	

#### Refinement

Refinement on $F$	$(\Delta/\sigma)_{\text{max}} = 0.69$
$R = 0.029$	$\Delta\rho_{\text{max}} = 0.15 \text{ e \AA}^{-3}$
$wR = 0.032$	$\Delta\rho_{\text{min}} = -0.14 \text{ e \AA}^{-3}$
$S = 1.139$	Extinction correction:
1042 reflections	Larson (1967)
227 parameters	Extinction coefficient:
All H-atom parameters refined	$2.6$ (3) $\times 10^5$

Table 1. Fractional atomic coordinates and equivalent isotropic displacement parameters ( $\text{\AA}^2$ ) for (A)

	$x$	$y$	$z$	$U_{\text{eq}}$
O(1)	0.53141 (6)	0.1144 (1)	0.65034	0.0584 (9)
O(2)	0.58115 (5)	0.0987 (1)	0.7767 (6)	0.0481 (7)
O(3)	0.62173 (5)	0.0707 (1)	0.3496 (6)	0.0471 (7)
N(1)	0.60952 (5)	0.1753 (1)	0.4827 (6)	0.0334 (7)
N(2)	0.56185 (7)	0.2324 (1)	0.1552 (6)	0.0453 (9)
C(1)	0.55988 (7)	0.1248 (1)	0.6385 (7)	0.0365 (9)
C(2)	0.57532 (7)	0.1666 (1)	0.4758 (7)	0.0344 (8)
C(3)	0.55590 (7)	0.1955 (1)	0.3296 (7)	0.0361 (9)
C(4)	0.5351 (1)	0.2603 (2)	0.0355 (9)	0.065 (2)
C(5)	0.5940 (1)	0.2447 (2)	0.0717 (8)	0.057 (1)
C(6)	0.5679 (1)	0.0561 (2)	0.9405 (8)	0.061 (1)
C(7)	0.63042 (7)	0.1258 (1)	0.4241 (6)	0.0332 (8)

C(71)	0.66549 (7)	0.1429 (1)	0.4490 (6)	0.0352 (8)
C(72)	0.67718 (8)	0.1820 (2)	0.6179 (8)	0.043 (1)
C(73)	0.70987 (8)	0.1951 (2)	0.6327 (8)	0.052 (1)
C(74)	0.73067 (7)	0.1710 (2)	0.4844 (9)	0.0550 (1)
C(75)	0.71939 (9)	0.1334 (2)	0.3150 (8)	0.0550 (1)
C(76)	0.68685 (8)	0.1183 (2)	0.2996 (8)	0.0460 (1)

Table 2. Selected geometric parameters ( $\text{\AA}$ ,  $^\circ$ ) for (A)

O(1)–C(1)	1.202 (4)	C(1)–C(2)	1.460 (5)
O(2)–C(1)	1.343 (5)	C(2)–C(3)	1.350 (5)
O(2)–C(6)	1.438 (6)	C(7)–C(71)	1.501 (4)
O(3)–C(7)	1.226 (4)	C(71)–C(76)	1.382 (5)
N(1)–C(7)	1.348 (4)	C(71)–C(72)	1.400 (5)
N(1)–C(2)	1.430 (4)	C(72)–C(73)	1.384 (5)
N(2)–C(3)	1.341 (5)	C(73)–C(74)	1.359 (6)
N(2)–C(4)	1.449 (6)	C(74)–C(75)	1.382 (7)
N(2)–C(5)	1.454 (5)	C(75)–C(76)	1.385 (5)
C(1)–O(2)–C(6)	115.9 (3)	O(3)–C(7)–N(1)	122.8 (3)
C(7)–N(1)–C(2)	123.1 (2)	O(3)–C(7)–C(71)	121.3 (3)
C(3)–N(2)–C(4)	119.4 (3)	N(1)–C(7)–C(71)	115.9 (2)
C(3)–N(2)–C(5)	123.8 (3)	C(76)–C(71)–C(72)	119.3 (3)
C(4)–N(2)–C(5)	116.7 (4)	C(76)–C(71)–C(7)	118.3 (3)
O(1)–C(1)–O(2)	122.8 (3)	C(72)–C(71)–C(7)	122.4 (3)
O(1)–C(1)–C(2)	124.7 (3)	C(73)–C(72)–C(71)	119.5 (4)
O(2)–C(1)–C(2)	112.5 (3)	C(74)–C(73)–C(72)	120.8 (4)
C(3)–C(2)–N(1)	124.3 (3)	C(73)–C(74)–C(75)	120.3 (3)
C(3)–C(2)–C(1)	117.0 (3)	C(74)–C(75)–C(76)	119.8 (4)
N(1)–C(2)–C(1)	118.7 (3)	C(71)–C(76)–C(75)	120.3 (4)
N(2)–C(3)–C(2)	132.7 (3)		

**Compound (B)***Crystal data* $M_r = 326.36$ 

Monoclinic

*Cc* $a = 36.050 (5) \text{ \AA}$  $b = 4.943 (1) \text{ \AA}$  $c = 19.793 (3) \text{ \AA}$  $\beta = 108.72 (1)^\circ$  $V = 3340 (2) \text{ \AA}^3$  $Z = 8$  $D_x = 1.298 \text{ Mg m}^{-3}$ *Data collection*

Enraf-Nonius CAD-4 diffractometer

 $\omega-2\theta$  scansAbsorption correction:  
none

8890 measured reflections

4454 independent reflections

1387 observed reflections

[ $I > 2.5\sigma(I)$ ]*Refinement*Refinement on  $F$  $R = 0.038$  $wR = 0.048$  $S = 1.130$ 

2317 reflections

290 parameters

All H-atom parameters refined

Mo  $K\alpha$  radiation $\lambda = 0.71069 \text{ \AA}$ 

Cell parameters from 100

reflections

 $\theta = 7.2-12.4^\circ$  $\mu = 0.856 \text{ mm}^{-1}$  $T = 293 \text{ K}$ 

Prismatic

0.6 × 0.3 × 0.2 mm

Colourless

R<sub>int</sub> = 0.045 $\theta_{\max} = 28^\circ$  $h = -47 \rightarrow 47$  $k = -6 \rightarrow 0$  $l = -26 \rightarrow 26$ 

3 standard reflections

frequency: 166.67 min

intensity variation: 3.8%

 $(\Delta/\sigma)_{\max} = 0.86$  $\Delta\rho_{\max} = 0.24 \text{ e \AA}^{-3}$  $\Delta\rho_{\min} = -0.25 \text{ e \AA}^{-3}$ 

Extinction correction:

Larson (1967)

Extinction coefficient:

 $1.94 (7) \times 10^5$ Table 3. Fractional atomic coordinates and equivalent isotropic displacement parameters ( $\text{\AA}^2$ ) for (B)

	$x$	$y$	$z$	$U_{\text{eq}}$
C(1)	0.07253 (8)	-0.0017 (5)	0.6469 (1)	0.0494 (9)
O(1)	0.07417 (7)	-0.1088 (5)	0.7019 (1)	0.0749 (9)
O(2)	0.04308 (6)	-0.0465 (5)	0.5860 (1)	0.0726 (8)
C(2)	0.10112 (7)	0.1918 (5)	0.6367 (1)	0.0400 (7)
C(3)	0.09432 (7)	0.3329 (5)	0.5766 (1)	0.0418 (7)
N(1)	0.11784 (6)	0.5293 (4)	0.56494 (9)	0.0448 (7)
C(4)	0.11205 (7)	0.6621 (5)	0.5005 (1)	0.0417 (8)
N(2)	0.08234 (6)	0.5822 (5)	0.4446 (1)	0.0506 (7)
C(5)	0.07962 (8)	0.7083 (6)	0.3828 (1)	0.0563 (9)
C(6)	0.10588 (8)	0.9053 (6)	0.3790 (1)	0.0556 (9)
C(7)	0.13538 (7)	0.9749 (5)	0.4412 (1)	0.0468 (8)
N(3)	0.13870 (6)	0.8515 (4)	0.5028 (1)	0.0453 (7)
C(8)	0.16535 (9)	1.1849 (6)	0.4424 (2)	0.062 (1)
C(9)	0.0472 (1)	0.6161 (9)	0.3181 (2)	0.093 (1)
N(4)	0.13563 (6)	0.2308 (4)	0.69611 (9)	0.0417 (6)
C(10)	0.16119 (7)	0.0286 (4)	0.7235 (1)	0.0403 (8)
O(3)	0.15630 (6)	-0.2020 (3)	0.69879 (9)	0.0552 (6)
C(11)	0.19622 (7)	0.0928 (5)	0.7859 (1)	0.0385 (7)
C(12)	0.22914 (8)	-0.0678 (6)	0.7973 (1)	0.0524 (9)
C(13)	0.26262 (8)	-0.0195 (7)	0.8541 (2)	0.063 (1)
C(14)	0.26311 (8)	0.1856 (6)	0.9011 (1)	0.061 (1)
C(15)	0.23056 (9)	0.3441 (6)	0.8915 (1)	0.062 (1)
C(16)	0.19707 (8)	0.3008 (5)	0.8334 (1)	0.0498 (8)
C(17)	0.0123 (11)	-0.223 (1)	0.5914 (3)	0.097 (2)

Table 4. Selected geometric parameters ( $\text{\AA}$ ,  $^\circ$ ) for (B)

C(1)–O(1)	1.195 (3)	C(6)–C(7)	1.387 (3)
C(1)–O(2)	1.344 (3)	C(7)–N(3)	1.334 (3)
C(1)–C(2)	1.467 (4)	C(7)–C(8)	1.493 (4)
O(2)–C(17)	1.442 (5)	N(4)–C(10)	1.350 (3)
C(2)–C(3)	1.332 (3)	C(10)–O(3)	1.231 (2)
C(2)–N(4)	1.424 (3)	C(10)–C(11)	1.490 (3)
C(3)–N(1)	1.357 (3)	C(11)–C(12)	1.385 (4)
N(1)–C(4)	1.389 (3)	C(11)–C(16)	1.388 (3)
C(4)–N(2)	1.329 (3)	C(12)–C(13)	1.381 (4)
C(4)–N(3)	1.332 (3)	C(13)–C(14)	1.372 (5)
N(2)–C(5)	1.348 (3)	C(14)–C(15)	1.372 (4)
C(5)–C(6)	1.377 (4)	C(15)–C(16)	1.391 (3)
C(5)–C(9)	1.501 (4)		
O(1)–C(1)–O(2)	122.5 (3)	N(3)–C(7)–C(6)	121.1 (2)
O(1)–C(1)–C(2)	125.6 (2)	N(3)–C(7)–C(8)	116.8 (2)
O(2)–C(1)–C(2)	111.8 (2)	C(6)–C(7)–C(8)	122.1 (3)
C(1)–O(2)–C(17)	116.0 (3)	C(4)–N(3)–C(7)	116.0 (2)
C(3)–C(2)–N(4)	121.9 (2)	C(10)–N(4)–C(2)	122.3 (2)
C(3)–C(2)–C(1)	121.9 (2)	O(3)–C(10)–N(4)	122.3 (2)
N(4)–C(2)–C(1)	116.2 (2)	O(3)–C(10)–C(11)	120.1 (2)
C(2)–C(3)–N(1)	125.5 (2)	N(4)–C(10)–C(11)	117.6 (2)
C(3)–N(1)–C(4)	125.2 (2)	C(12)–C(11)–C(16)	118.9 (2)
N(2)–C(4)–N(3)	128.1 (2)	C(12)–C(11)–C(10)	117.8 (2)
N(2)–C(4)–N(1)	117.9 (2)	C(16)–C(11)–C(10)	123.2 (2)
N(3)–C(4)–N(1)	114.0 (2)	C(13)–C(12)–C(11)	120.9 (3)
C(4)–N(2)–C(5)	115.0 (2)	C(14)–C(13)–C(12)	119.8 (3)
N(2)–C(5)–C(6)	121.7 (2)	C(13)–C(14)–C(15)	120.2 (2)
N(2)–C(5)–C(9)	116.3 (3)	C(14)–C(15)–C(16)	120.2 (3)
C(6)–C(5)–C(9)	121.9 (3)	C(11)–C(16)–C(15)	119.9 (3)
C(5)–C(6)–C(7)	118.1 (2)		

**Compound (C)***Crystal data* $M_r = 427.46$ 

Monoclinic

*P2<sub>1</sub>* $a = 9.398 (1) \text{ \AA}$  $b = 18.422 (2) \text{ \AA}$  $c = 12.663 (1) \text{ \AA}$  $\beta = 91.46 (1)^\circ$ Mo  $K\alpha$  radiation $\lambda = 0.71069 \text{ \AA}$ 

Cell parameters from 75

reflections

 $\theta = 8.07-12.65^\circ$  $\mu = 0.883 \text{ mm}^{-1}$  $T = 293 \text{ K}$ 

Prismatic

$V = 2191.8 (7) \text{ \AA}^3$  $Z = 4$  $D_x = 1.296 \text{ Mg m}^{-3}$  $D_m = 1.289 \text{ Mg m}^{-3}$  $D_m$  measured by flotation**Data collection**

Enraf-Nonius CAD-4

diffractometer

 $\omega$ - $2\theta$  scans

Absorption correction:

none

10 173 measured reflections

5049 independent reflections

2404 observed reflections

[ $I > 2.5\sigma(I)$ ]**Refinement**Refinement on  $F$  $R = 0.034$  $wR = 0.041$  $S = 0.7469$ 

3617 reflections

558 parameters

 $0.76 \times 0.32 \times 0.20 \text{ mm}$ 

Colourless

 $R_{\text{int}} = 0.015$  $\theta_{\text{max}} = 28^\circ$  $h = -12 \rightarrow 12$  $k = -24 \rightarrow 24$  $l = 0 \rightarrow 16$ 

3 standard reflections

frequency: 200 min

intensity variation: 0.92%

C(7B)	-0.0388 (3)	0.3843 (2)	0.6376 (3)	0.051 (1)
C(8B)	-0.3265 (3)	0.4289 (2)	0.7970 (3)	0.050 (1)
O(2B)	-0.2904 (3)	0.4944 (2)	0.7669 (2)	0.0641 (9)
O(3B)	-0.4353 (3)	0.4150 (2)	0.8418 (3)	0.068 (1)
N(2B)	0.2355 (2)	0.3873 (2)	0.7639 (2)	0.0402 (8)
C(9B)	0.2800 (3)	0.3272 (2)	0.8157 (3)	0.043 (1)
O(4B)	0.2510 (3)	0.3155 (1)	0.9083 (2)	0.0580 (8)
C(10B)	0.3643 (3)	0.2740 (2)	0.7549 (3)	0.0428 (9)
C(11B)	0.3494 (3)	0.2657 (2)	0.6464 (3)	0.050 (1)
C(12B)	0.4251 (4)	0.2120 (2)	0.5951 (3)	0.059 (1)
C(13B)	0.5180 (4)	0.1681 (2)	0.6501 (4)	0.073 (2)
C(14B)	0.5348 (5)	0.1765 (3)	0.7586 (4)	0.073 (2)
C(15B)	0.4587 (4)	0.2285 (2)	0.8107 (3)	0.062 (1)
N(3B)	0.3391 (3)	0.5224 (2)	0.8406 (3)	0.056 (1)
C(16B)	0.3907 (4)	0.5899 (2)	0.8633 (3)	0.049 (1)
N(5B)	0.3008 (4)	0.6400 (2)	0.8962 (2)	0.059 (1)
C(19B)	0.3565 (6)	0.7058 (2)	0.9174 (3)	0.068 (2)
C(18B)	0.4978 (6)	0.7196 (3)	0.9044 (3)	0.076 (2)
C(17B)	0.5838 (5)	0.6644 (2)	0.8709 (3)	0.063 (1)
N(4B)	0.5306 (3)	0.5984 (2)	0.8489 (2)	0.055 (1)
C(21B)	0.2547 (8)	0.7617 (3)	0.9573 (4)	0.097 (2)
C(20B)	0.7423 (6)	0.6725 (3)	0.8586 (4)	0.091 (2)
O(20B)	0.1061 (3)	0.3802 (2)	1.0801 (2)	0.077 (1)

Table 6. Selected geometric parameters ( $\text{\AA}$ ,  $^\circ$ ) for (C)

C(1A)—C(2A)	1.325 (5)	C(1B)—C(2B)	1.317 (5)
C(1A)—N(3A)	1.389 (4)	C(1B)—N(3B)	1.392 (4)
C(2A)—N(2A)	1.431 (4)	C(2B)—N(2B)	1.431 (4)
C(2A)—C(3A)	1.496 (4)	C(2B)—C(3B)	1.494 (4)
C(3A)—O(1A)	1.233 (4)	C(3B)—O(1B)	1.235 (4)
C(3A)—N(1A)	1.337 (4)	C(3B)—N(1B)	1.338 (4)
N(1A)—C(4A)	1.459 (4)	N(1B)—C(7B)	1.474 (4)
N(1A)—C(7A)	1.470 (4)	N(1B)—C(4B)	1.479 (4)
C(4A)—C(8A)	1.509 (5)	C(4B)—C(8B)	1.511 (5)
C(4A)—C(5A)	1.529 (7)	C(4B)—C(5B)	1.535 (6)
C(5A)—C(6A)	1.501 (7)	C(5B)—C(6B)	1.430 (7)
C(6A)—C(7A)	1.509 (6)	C(6B)—C(7B)	1.490 (6)
C(8A)—O(3A)	1.214 (4)	C(8B)—O(3B)	1.209 (4)
C(8A)—O(2A)	1.314 (5)	C(8B)—O(2B)	1.312 (5)
N(2A)—C(9A)	1.341 (4)	N(2B)—C(9B)	1.346 (4)
C(9A)—O(4A)	1.235 (4)	C(9B)—O(4B)	1.230 (4)
C(9A)—C(10A)	1.493 (4)	C(9B)—C(10B)	1.488 (5)
C(10A)—C(11A)	1.378 (5)	C(10B)—C(11B)	1.385 (5)
C(10A)—C(15A)	1.384 (5)	C(10B)—C(15B)	1.399 (5)
C(11A)—C(12A)	1.388 (6)	C(11B)—C(12B)	1.389 (5)
C(12A)—C(13A)	1.374 (7)	C(12B)—C(13B)	1.386 (6)
C(13A)—C(14A)	1.360 (6)	C(13B)—C(14B)	1.388 (7)
C(14A)—C(15A)	1.387 (6)	C(14B)—C(15B)	1.375 (6)
N(3A)—C(16A)	1.376 (4)	N(3B)—C(16B)	1.363 (5)
C(16A)—N(5A)	1.330 (4)	C(16B)—N(5B)	1.326 (5)
C(16A)—N(4A)	1.347 (4)	C(16B)—N(4B)	1.342 (4)
N(5A)—C(19A)	1.349 (5)	N(5B)—C(19B)	1.344 (6)
C(19A)—C(18A)	1.371 (7)	C(19B)—C(18B)	1.366 (8)
C(19A)—C(21A)	1.501 (7)	C(19B)—C(21B)	1.501 (8)
C(18A)—C(17A)	1.362 (6)	C(18B)—C(17B)	1.374 (7)
C(17A)—N(4A)	1.337 (5)	C(17B)—N(4B)	1.340 (5)
C(17A)—C(20A)	1.499 (6)	C(17B)—C(20B)	1.509 (7)
C(2A)—C(1A)—N(3A)	121.9 (3)	C(2B)—C(1B)—N(3B)	123.4 (3)
C(1A)—C(2A)—N(2A)	121.1 (3)	C(1B)—C(2B)—N(2B)	121.7 (3)
C(1A)—C(2A)—C(3A)	120.7 (3)	C(1B)—C(2B)—C(3B)	119.0 (3)
N(2A)—C(2A)—C(3A)	118.2 (3)	N(2B)—C(2B)—C(3B)	119.0 (3)
O(1A)—C(3A)—C(2A)	121.8 (3)	O(1B)—C(3B)—N(1B)	120.8 (3)
O(1A)—C(3A)—N(1A)	121.8 (3)	O(1B)—C(3B)—C(2B)	121.3 (3)
C(3A)—N(1A)—C(2A)	118.2 (3)	N(1B)—C(3B)—C(2B)	117.9 (3)
C(3A)—N(1A)—C(4A)	118.9 (3)	C(3B)—N(1B)—C(7B)	128.2 (3)
O(4A)—C(4A)—N(1A)	128.4 (3)	C(3B)—N(1B)—C(4B)	119.1 (3)
C(4A)—N(1A)—C(7A)	112.1 (3)	C(7B)—N(1B)—C(4B)	112.1 (2)
C(14A)—C(1A)—C(2A)	114.4 (3)	N(1B)—C(4B)—C(8B)	114.5 (3)
C(14A)—C(1A)—N(1A)	103.6 (3)	N(1B)—C(4B)—C(5B)	102.7 (3)
N(1A)—C(4A)—C(5A)	110.8 (3)	C(8B)—C(4B)—C(5B)	112.7 (3)
C(8A)—C(4A)—C(5A)	103.7 (4)	C(6B)—C(5B)—C(4B)	106.1 (4)
C(6A)—C(5A)—C(4A)	105.2 (4)	C(5B)—C(6B)—C(7B)	109.5 (4)
C(5A)—C(6A)—C(7A)	103.4 (3)	N(1B)—C(7B)—C(6B)	102.9 (3)
N(1A)—C(7A)—C(6A)	123.3 (3)	O(3B)—C(8B)—O(2B)	124.0 (3)
O(3A)—C(8A)—O(2A)	120.8 (3)	O(3B)—C(8B)—C(4B)	119.9 (3)
O(3A)—C(8A)—C(4A)	115.9 (3)	O(2B)—C(8B)—C(4B)	115.9 (3)
C(9A)—N(2A)—C(2A)	120.4 (3)	C(9B)—N(2B)—C(2B)	121.4 (3)

Table 5. Fractional atomic coordinates and equivalent isotropic displacement parameters ( $\text{\AA}^2$ ) for (C)

$U_{\text{eq}} = (1/3)\sum_i U_{ij}a_i^*a_j^*$	$\mathbf{a}_i \cdot \mathbf{a}_j$	Atoms of molecules (I) and (II) of the asymmetric unit are indicated by the labels A and B, respectively.
C(1A)	0.2639 (3)	0.3349 (2)
C(2A)	0.3153 (3)	0.3973 (2)
C(3A)	0.4686 (3)	0.4164 (2)
O(1A)	0.5288 (2)	0.40030
N(1A)	0.5361 (3)	0.4518 (2)
C(4A)	0.6810 (3)	0.4772 (2)
C(5A)	0.7174 (4)	0.5182 (3)
C(6A)	0.6289 (5)	0.4812 (3)
C(7A)	0.4923 (4)	0.4607 (2)
C(8A)	0.7876 (3)	0.4175 (2)
O(2A)	0.7491 (3)	0.3521 (2)
O(3A)	0.9026 (2)	0.4309 (2)
N(2A)	0.2252 (2)	0.4494 (1)
C(9A)	0.2021 (3)	0.5144 (2)
O(4A)	0.2483 (3)	0.5302 (1)
C(10A)	0.1198 (3)	0.5689 (2)
C(11A)	0.0604 (4)	0.6268 (2)
C(12A)	-0.0150 (5)	0.6797 (3)
C(13A)	-0.0301 (4)	0.6748 (2)
C(14A)	0.0301 (4)	0.6183 (2)
C(15A)	0.1053 (4)	0.5650 (2)
N(3A)	0.1197 (3)	0.3186 (2)
C(16A)	0.0616 (4)	0.2526 (2)
N(5A)	0.1465 (3)	0.1997 (2)
C(19A)	0.0822 (5)	0.1365 (2)
C(18A)	-0.0627 (5)	0.1290 (2)
C(17A)	-0.1425 (4)	0.1863 (2)
N(4A)	-0.0809 (3)	0.2488 (2)
C(21A)	0.1788 (7)	0.0775 (3)
C(20A)	-0.3020 (5)	0.1848 (3)
O(10A)	0.3496 (4)	0.4490 (2)
C(1B)	0.1966 (3)	0.5025 (2)
C(2B)	0.1464 (3)	0.4397 (2)
C(3B)	-0.0066 (3)	0.4217 (2)
O(1B)	-0.0645 (2)	0.4347 (2)
N(1B)	-0.0796 (3)	0.3907 (2)
C(4B)	-0.2286 (3)	0.3682 (2)
C(5B)	-0.2715 (5)	0.3353 (3)
C(6B)	-0.1757 (5)	0.3651 (4)

O(4A)—C(9A)—N(2A)	122.3 (3)	O(4B)—C(9B)—N(2B)	122.3 (3)
O(4A)—C(9A)—C(10A)	120.1 (3)	O(4B)—C(9B)—C(10B)	120.8 (3)
N(2A)—C(9A)—C(10A)	117.5 (3)	N(2B)—C(9B)—C(10B)	116.9 (3)
C(11A)—C(10A)—C(15A)	118.8 (3)	C(11B)—C(10B)—C(15B)	118.8 (3)
C(11A)—C(10A)—C(9A)	118.3 (3)	C(11B)—C(10B)—C(9B)	122.9 (3)
C(15A)—C(10A)—C(9A)	122.9 (3)	C(15B)—C(10B)—C(9B)	118.2 (3)
C(10A)—C(11A)—C(12A)	120.5 (4)	C(10B)—C(11B)—C(12B)	120.1 (3)
C(13A)—C(12A)—C(11A)	120.1 (4)	C(13B)—C(12B)—C(11B)	120.7 (4)
C(14A)—C(13A)—C(12A)	119.7 (4)	C(12B)—C(13B)—C(14B)	119.6 (4)
C(13A)—C(14A)—C(15A)	120.8 (4)	C(15B)—C(14B)—C(13B)	120.3 (4)
C(10A)—C(15A)—C(14A)	120.1 (3)	C(14B)—C(15B)—C(10B)	120.4 (4)
C(16A)—N(3A)—C(1A)	124.6 (3)	C(16B)—N(3B)—C(1B)	124.4 (3)
N(5A)—C(16A)—N(4A)	126.1 (3)	N(5B)—C(16B)—N(4B)	126.6 (3)
N(5A)—C(16A)—N(3A)	119.3 (3)	N(5B)—C(16B)—N(3B)	118.4 (3)
N(4A)—C(16A)—N(3A)	114.6 (3)	N(4B)—C(16B)—N(3B)	115.0 (3)
C(16A)—N(5A)—C(19A)	116.0 (3)	C(16B)—N(5B)—C(19B)	116.3 (4)
N(5A)—C(19A)—C(18A)	121.1 (4)	N(5B)—C(19B)—C(18B)	121.2 (4)
N(5A)—C(19A)—C(21A)	115.6 (4)	N(5B)—C(19B)—C(21B)	115.9 (5)
C(18A)—C(19A)—C(21A)	123.3 (4)	C(18B)—C(19B)—C(21B)	122.9 (4)
C(17A)—C(18A)—C(19A)	119.3 (4)	C(19B)—C(18B)—C(17B)	118.8 (4)
N(4A)—C(17A)—C(18A)	120.8 (4)	N(4B)—C(17B)—C(18B)	121.1 (4)
N(4A)—C(17A)—C(20A)	116.4 (4)	N(4B)—C(17B)—C(20B)	115.6 (4)
C(18A)—C(17A)—C(20A)	122.7 (4)	C(18B)—C(17B)—C(20B)	123.3 (5)
C(17A)—N(4A)—C(16A)	116.7 (3)	C(17B)—N(4B)—C(16B)	116.1 (3)

Table 7. Hydrogen-bond distances ( $\text{\AA}$ ) between water molecules and molecules (I) and (II) of the asymmetric unit of compound (C)

O(10)···O(1A)	2.822 (4)	O(20)···O(1B)	2.804 (4)
O(10)···N(2B)	2.846 (4)	O(20)···O(2A <sup>1</sup> )	2.835 (4)
O(10)···O(4A)	2.902 (4)	O(20)···O(4A)	2.856 (4)

Symmetry code: (i)  $x, y, z - 1$ .

For compound (A):  $w = W_f W_s$ , where  $W_f(|F_o| < 25) = (|F_o|/25)^{1.5}$ ,  $W_f(|F_o| > 33) = (33/|F_o|)^{0.5}$ ,  $W_f(25 < |F_o| < 33) = 1.0$ ,  $W_s(\sin\theta < 0.30) = (\sin\theta/0.30)^{3.0}$ ,  $W_s(\sin\theta > 0.32) = (0.32/\sin\theta)^{3.0}$ ,  $W_s(0.30 < \sin\theta < 0.30) = 1.0$ .

For compound (B):  $w = 2W_f W_s$ , where  $W_f(|F_o| < 19) = (|F_o|/19)^{1.0}$ ,  $W_f(|F_o| > 26) = (33/|F_o|)^{1.5}$ ,  $W_f(19 < |F_o| < 26) = 1.0$ ,  $W_s(\sin\theta < 0.36) = (\sin\theta/0.36)^{1.5}$ ,  $W_s(\sin\theta > 0.39) = (0.39/\sin\theta)^{5.0}$ ,  $W_s(0.36 < \sin\theta < 0.39) = 1.0$ .

For compound (C):  $w = 4W_f W_s$ , where  $W_f(|F_o| < 8) = (|F_o|/8)^{1.0}$ ,  $W_f(|F_o| > 14) = (14/|F_o|/25)^{1.0}$ ,  $W_f(8 < |F_o| < 14) = 1.0$ ,  $W_s(\sin\theta < 0.33) = (\sin\theta/0.33)^{2.0}$ ,  $W_s(\sin\theta > 0.37) = (0.37/\sin\theta)^{6.0}$ ,  $W_s(0.33 < \sin\theta < 0.37) = 1.0$ ; the positions of H atoms were calculated and included in structure-factor calculations as fixed-atom contributions.

For all compounds, data collection: Enraf–Nonius CAD-4 software. Cell refinement: XRAY76 PARAM (Stewart, Machin, Dickinson, Ammon, Heck & Flack, 1976). Data reduction: XRAY76 DATCOR, DATRDN. Program(s) used to solve structure: MULTAN80 (Main, Fiske, Hull, Lessinger, Germain, Declercq & Woolfson, 1980) for (A), (B); SHELXS86 (Sheldrick, 1985) for (C). Program(s) used to refine structure: XRAY76 CRYSLQ. Software used to prepare material for publication: XRAY76 BONDLA, LISTFC; Xtal3.0 BONDLA (Hall & Stewart, 1990). Scattering factors for neutral atoms from Cromer & Mann (1968), for H atoms from Stewart, Davidson & Simpson (1965).

Lists of structure factors, anisotropic displacement parameters, H-atom coordinates, complete geometry, least-squares-planes data and torsion angles have been deposited with the British Library Document Supply Centre as Supplementary Publication No. SUP 71458 (52 pp.). Copies may be obtained through The Technical Editor, International Union of Crystallography, 5 Abbey Square, Chester CH1 2HU, England. [CIF reference: NA1033]

## References

- Allen, F. H., Kennard, O., Watson, D. G., Brammer, L., Orpen, A. G. & Taylor, R. (1987). *J. Chem. Soc. Perkin Trans. 2*, pp. S1–S19.
- Ashida, T. & Kakudo, M. (1974). *Bull. Chem. Soc. Jpn.*, **47**, 1129–1133.
- Balasubramanian, R., Lakshminarayanan, A. V., Sabesan, M. N., Tegoni, G., Venkatesan, K. & Ramachandran, G. N. (1971). *Int. J. Protein Res.*, **3**, 25–33.
- Cromer, D. T. & Mann, J. B. (1968). *Acta Cryst. A*, **24**, 321–324.
- El-Baba, S., Nuzillard, J. M., Paulin, J. C. & Kagan, H. B. (1986). *Tetrahedron*, **42**, 3851–3861.
- Epp, O., Lattman, E. E., Schiffer, M., Huber, R. & Palm, W. (1975). *Biochemistry*, **14**, 4943–4952.
- Hall, S. R. & Stewart, J. M. (1990). Editors. *Xtal3.0 Reference Manual*. Univs. of Western Australia, Australia, and Maryland, USA.
- Japan Kokai (1975). Jpn. patent 75/58 063; *Chem. Abstr.*, **83**, P193075y.
- Johnson, C. K. (1965). *ORTEP*. Report ORNL-3784. Oak Ridge National Laboratory, Tennessee, USA.
- Kagan, H. B. (1985). In *Asymmetric Synthesis*, Vol. 5, edited by J. D. Morrison. Orlando: Academic.
- Kmetič, M., Stanovnik, B. & Tišler, M. (1993). *Heterocycles*, **35**, 1331–1339.
- Larson, A. C. (1967). *Acta Cryst.*, **23**, 664–665.
- Leban, I., Sveti, J., Stanovnik, B. & Tišler, M. (1991). *Acta Cryst. C*, **47**, 1552–1554.
- Main, P., Fiske, S. J., Hull, S. E., Lessinger, L., Germain, G., Declercq, J.-P. & Woolfson, M. M. (1980). *MULTAN80. A System of Computer Programs for the Automatic Solution of Crystal Structures from X-ray Diffraction Data*. Univs. of York, England, and Louvain, Belgium.
- Ornik, B., Čadež, Z., Stanovnik, B. & Tišler, M. (1990). *J. Heterocycl. Chem.*, **27**, 1021–1024.
- Ornik, B., Stanovnik, B. & Tišler, M. (1992a). *J. Heterocycl. Chem.*, **29**, 831–834.
- Ornik, B., Stanovnik, B. & Tišler, M. (1992b). *J. Heterocycl. Chem.*, **29**, 1241–1244.
- Pauling, L. (1960). *The Nature of the Chemical Bond*, 3rd ed., pp. 281. Ithaca: Cornell Univ. Press.
- Ramachandran, G. N. & Ramakrishnan, C. (1965). *Biophys. J.*, **5**, 909–933.
- Ramachandran, G. N., Ramakrishnan, C. & Sasisekharan, V. (1963). *J. Mol. Biol.*, **7**, 95–99.
- Schmidt, U., Häusler, J., Öhler, E. & Poisel, H. (1979). *Dehydro-amino Acids, α-Hydroxy-α-amino Acids and α-Mercapto-α-amino Acids. In Progress in the Chemistry of Organic Natural Products*, Vol. 37, edited by W. Herz, H. Grisebach & G. W. Kirby, pp. 215–327. Vienna: Springer.
- Schmidt, U., Lieberknecht, A. & Wild, J. (1988). *Synthesis*, pp. 159–172.
- Sheldrick, G. M. (1985). *SHELXS86. Crystallographic Computing 3*, edited by G. M. Sheldrick, C. Krüger & R. Goddard, pp. 175–189. Oxford Univ. Press.
- Stammer, C. H. (1982). *Chemistry and Biochemistry of Amino Acids, Peptides and Proteins*, Vol. 6, pp. 33–74. London: John Wiley.
- Stanovnik, B., Golič, L., Kmecl, P., Ornik, B., Sveti, J. & Tišler, M. (1991). *J. Heterocycl. Chem.*, **28**, 1961–1964.
- Stanovnik, B., Sveti, J. & Tišler, M. (1987). *J. Heterocycl. Chem.*, **24**, 1809–1810.
- Stanovnik, B., Sveti, J. & Tišler, M. (1989). *J. Heterocycl. Chem.*, **26**, 1273–1275.
- Stanovnik, B., Sveti, J., Tišler, M., Žorž, L., Hvala, A. & Simonič, I. (1988). *Heterocycles*, **27**, 903–909.
- Stanovnik, B., Urbanija, M., Sveti, J. & Tišler, M. (1989). *Arch. Pharm. (Weinheim)*, **322**, 783–787.

- Stanovnik, B., van de Bovenkamp, H., Svetec, J., Hvala, A., Simonić, I. & Tišler, M. (1989). *J. Heterocycl. Chem.* **27**, 359–361.
- Stewart, J. M., Machin, P. A., Dickinson, C. W., Ammon, H. L., Heck, H. & Flack, H. (1976). *The XRAY76 System*. Technical Report TR-446. Computer Science Center, Univ. of Maryland, College Park, Maryland, USA.
- Stewart, R. F., Davidson, E. R. & Simpson, W. T. (1965). *J. Chem. Phys.* **42**, 3175–3187.
- Svetec, J., Čadež, Z., Stanovnik, B. & Tišler, M. (1990). *Synthesis*, pp. 70–72.
- Svetec, J., Stanovnik, B., Tišler, M., Golič, L. & Leban, I. (1989). *J. Heterocycl. Chem.* **26**, 145–153.
- Wieland, T. & Birr, C. (1976). *MTP International Review of Science. Organic Chemistry, Series 2*, Vol. 6, edited by H. N. Rydon, pp. 183–218. London: Butterworths.

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## Double-Layered Polytypic Structure of the B Form of Octadecanoic Acid, $C_{18}H_{36}O_2$

FUMITOSHI KANEKO, HIROTOSHI SAKASHITA AND MASAMICHI KOBAYASHI

Department of Macromolecular Science,  
Faculty of Science, Osaka University, Toyonaka,  
Osaka 560, Japan

YASUYUKI KITAGAWA AND YOSHIKI MATSUURA

Institute for Protein Research, Osaka University, Suita,  
Osaka 565, Japan

MASAO SUZUKI

Oleochemicals Research Laboratory, Nippon Oil and  
Fats Co., Amagasaki 660, Japan

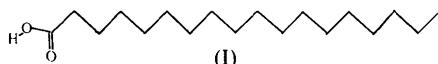
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### Abstract

Two bimolecular layers forming a double-layered polytypic structure exist in a repeating unit along the stacking direction. One bimolecular layer has essentially the same structure as that of the ordinary single-layered type of the B form.

### Comment

The structure determination of the orthorhombic modification of the B form of stearic acid (I) (octadecanoic acid) has been undertaken to confirm its



double-layered polytypic structure, which was indicated from a previous vibrational spectroscopic study. Stearic acid crystallizes in four polymorphic phases, A, B, C and E. In the previous vibrational spectroscopic work (Kobayashi, Kobayashi, Itoh & Sato, 1984), we found a new type of single crystal of the B form. This crystal exhibited essentially the same crystal morphology and vibrational spectra as those of the ordinary monoclinic modification (single-layered structure, referred as *Mon*) of the B form (Goto & Asada, 1978); however, large differences were observed in the low-frequency region of Raman spectra and in the bands due to the methyl groups. On the basis of the spectral features and the Weissenberg photograph, we concluded that this new crystal of the B form is an orthorhombic modification of the double-layered polytypic structure (referred as *Orth II*) that had been found in *n*-alkanes and *n*-alcohols (Amelinckz, 1955, 1956; Boistelle, Simon & Pepé, 1976; Kobayashi, Kobayashi, Itoh, Chatani & Tadokoro, 1980). The systematic differences in thermodynamic stability and mechanical properties between the *Mon* and *Orth II* types have been studied by Sato, Kobayashi & Morishita (1988) and Itoh & Kobayashi (1991).

The present structure analysis shows that the B form has a double-layered polytypic structure, as expected (Fig. 1). The *c* dimension [87.662 (9) Å] is twice the thickness of one bimolecular layer in the *Mon* type (43.87 Å), whose lattice parameters are *a* = 5.587, *b* = 7.386, *c* = 49.33 Å and  $\beta$  = 117.24°. The eight molecules in the unit cell form two bimolecular layers related to each other by a twofold *c*-screw axis operation. Thus, the acyl chain inclines from the *c* axis toward the [010] and [0̄10] directions alternately, forming a herringbone structure. The arrangement of the methyl groups at the interface of the bimolecular layers is markedly different from that in the ordinary single-layered structure. In addition to the orientation of the terminal methyl groups, there is a systematic difference between the *Orth II* and *Mon* types in the distances between neighboring terminal methyls. The *c*-axis projection of the arrangement of the terminal methyls is depicted in Fig. 2, in comparison with that of the *Mon* type. In the *Orth II* type, a methyl C atom in one layer is located at the center of a dimple surrounded by four methyl C atoms in the opposite layer, that is, the methyl C atoms are placed at equal intervals along both the *a*<sub>s</sub> and *b*<sub>s</sub> directions. In case of the *Mon* type, the methyl C atom is displaced from the center along the *b*<sub>s</sub> direction. The methyl terminals in the *Mon* and *Orth II* types are in different intermolecular force fields, giving rise to different vibrational frequencies of the methyl group.

One bimolecular layer in the *Orth II* type of the B form has essentially the same structure as that of the